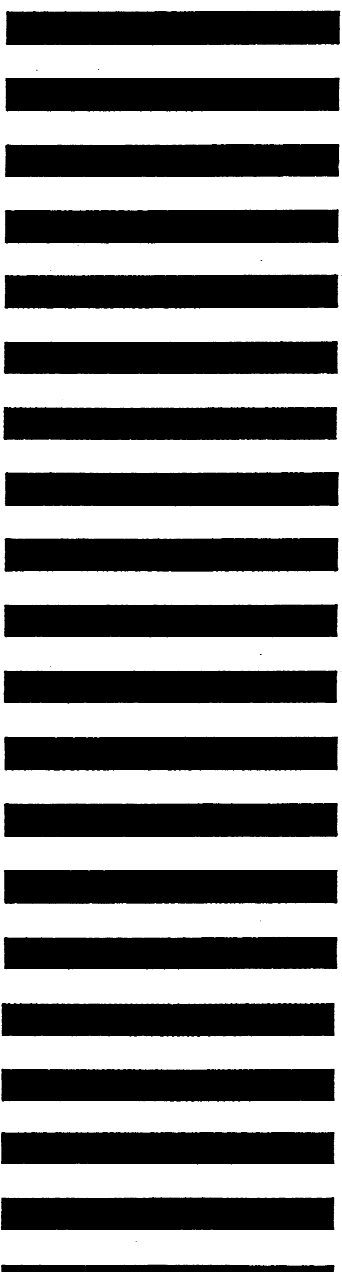




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INCLUDING ELECTROPHORESIS, MASS SPECTROMETRY
AND OTHER SEPARATION AND DETECTION METHODS

JOURNAL OF CHROMATOGRAPHY B

ANALYTICAL TECHNOLOGIES IN THE
BIOMEDICAL AND LIFE SCIENCES

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INSTRUCTIONS TO AUTHORS

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including electrophoresis, mass spectrometry and other separation and detection methods

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Journal of Chromatography B

analytical technologies in the biomedical and life sciences

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Figures should be submitted in a form suitable for reproduction, either drawn in Indian ink on drawing or tracing paper, or as sharp prints [either photographic (glossy) prints or prints from a high-resolution laser printer]. All axes of graphs and chromatograms should be clearly labelled, with full quantitative data, or equivalent information should be provided in the legend. Please note that any lettering should also be in a form suitable for reproduction. Lettering (which should be kept to a minimum) and spacing on axes of graphs should be such that numbers, etc., remain legible after reduction in size. One reproducible copy and three photocopies are required. The figures should preferably be of such a size that the same degree of reduction can be applied to all of them. The size of the figures should preferably not exceed the size of the text pages. Simple straight-line graphs (such as calibration lines) are not acceptable, because they can readily be described in the text by means of an equation or a sentence. Claims of linearity should be supported by regression data that include slope, intercept, standard deviations of the slope and intercept, standard error and the number of data points; correlation coefficients are optional. Standard symbols should be used in line drawings; the following are available to the typesetters and can also be used in the legends: filled or open squares, triangles, circles or diamonds, + or ×.

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[2] T.R. Bott. *Fouling of Heat Exchangers*. Elsevier, Amsterdam, 1995.

[3] C.H. Foyer, in R.G. Alscher, J.L. Hess (Editors), *Antioxidants in Higher Plants*. CRC Press, Boca Raton, FL, 1993, p. 31.

[4] A. Veide, C. Hassinen, D. Hallen, M. Eiteman, B. Lassen, K. Holmbert, in R.D. Rogers, M.A. Eiteman (Editors), *Proceedings of the American Chemical Society Symposium on Aqueous Biophasic Separation*. Plenum Publishers, New York, NY, 1995, p. 133.

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Appendix 1: Experimental conditions to be specified

Experimental conditions should preferably be given on a separate sheet, headed "Conditions". These conditions will, if appropriate, be printed in a block, directly following the heading "Experimental".

General

Chemicals. Supplier (+ city/town, state, country) and degree of purity of all less common chemicals; EC number of enzymes; optical purity of enantiomers.

Equipment. Model and manufacturer (+ city/town, state, country) of commercial instruments (e.g. chromatographs and detectors). For instruments that are not commercially available, sufficient detail (or a reference) should be given to allow others to construct their own instrument. Detection parameters (e.g. type, wavelength, attenuation, linearity range, limit of detection at a specified signal-to-noise ratio).

Sample preparation. Application papers should contain full details (or a reference) of the method of sample preparation. For centrifugation steps, give details of g value and time. Injection device and volume and concentration of the injected sample should be specified.

Column liquid chromatography

Column. Column dimensions (length \times internal diameter), manufacturer and location, packing material (for non-commercial columns or columns that are not widely used the chemical composition should be specified), particle diameter, pore diameter, column temperature.

Mobile phase. Complete and unambiguous description of the mobile phase composition or procedure for its preparation; pH; flow-rate; gradient programme.

k values. When reporting values, the method for determining the hold-up time (t_0) must be described.

Gas chromatography and supercritical fluid chromatography

Column. In addition to the parameters mentioned for column liquid chromatography, specify type of column (packed, capillary, etc.) support material, film thickness of the stationary phase, and surface modification, if applicable.

Carrier gas. Type, purity, flow-rate or inlet pressure (bar or MPa).

Temperature. All relevant temperatures (or temperature programmes) should be detailed.

Planar chromatography

Chamber. Internal dimensions, manufacturer and location, saturation, temperature, humidity.

Thin layer or paper. Manufacturer and location, material, dimensions, type (laboratory-prepared or commercially precoated) and thickness of layer, additives (fluorescent indicator, binder), position of starting line, development mode, method of activation.

Solvent. Composition of solvent, monophasic or upper or lower phase of two-phase mixture, total volume.

Sample. Application method, size of spot or streak, solvent and amount of solute and volume of solution applied.

Detection. Spray reagent, wavelength, details of colours, R_f values.

Electrophoresis

Matrix. For example, cellulose acetate, agarose, polyacrylamide; gel concentration; percentage cross-linker; dimensions and material of tube, sheet, etc., surface modification, length between column inlet and detector, temperature.

Buffers. Complete and unambiguous description of buffers used, pH and how the pH was set or adjusted.

Other. Injection method, voltage, current. In electropherograms, anode and cathode should be indicated.

Mass spectrometry

Inlet system. Direct on-line, off-line, postcolumn splitting, postcolumn buffer or matrix addition.

Source. Ionization energy, temperature, trap current, reagent gas. For LC interfaces, complete and unambiguous description of the same and their operating parameters (vaporizer and capillary temperature, buffers, nebulizing, auxiliary or ionizing gases, gas pressures, source and interface voltages, up-front CID voltages).

Mass analyzer. Accelerating voltage, scan mode, collision gas for tandem MS work, collision gas pressure, collision energy, resolution and mass range.

Detection. Electron multiplier voltage and/or electrometer gain, ions monitored in SIM and dwell times.

Appendix 2: Conversion table for the non-SI units most frequently used

The use of some non-SI units has been accepted for practical reasons; to this category belong units for time (min, h), volume (l), pressure (1 bar = 10^5 Pa), temperature ($^{\circ}\text{C}$), energy (1 eV \approx $160\,219 \cdot 10^{-21}$ J), mass (1 u \approx $1.66053 \cdot 10^{-27}$ kg) and activity (1 Ci = $3.7 \cdot 10^{10}$ Bq). This journal also accepts Å (= 0.1 nm). Concentration should formally be expressed in mol dm⁻³ or mol l⁻¹, but the symbol *M* is accepted; normality (*N*) should not be used, however. The frequently used “daltons” are not compatible with the SI system — the relative molecular mass (*M_r*) should be given as a value only (dimensionless). Gravitational force must be expressed in *g*; rpm is not allowed for centrifugation (but it is, e.g., for vortex mixing). The table below summarizes some conversion factors; to obtain the value in SI units, the value in non-SI units should be multiplied by the factor.

| Physical quantity | Type of conversion | Factor |
|-------------------|---------------------------------------|------------|
| Length | in. → cm | 2.54 |
| | ft. → cm | 30.4801 |
| Area | in. ² → cm ² | 6.451626 |
| Mass | lb. → kg | 0.45359237 |
| Volume | gallon (USA) → l | 3.785332 |
| | gallon (UK) → l | 4.54609 |
| Pressure | atm → Pa | 101 325 |
| | mmHg or Torr → Pa | 133.322 |
| | mmH ₂ O → Pa | 9.80665 |
| | kp cm ² → Pa | 98066.5 |
| | lbs. in. ⁻² or p.s.i. → Pa | 6894.76 |

Other frequently used non-SI “units” are ppm, ppb and ppt. When used in this journal, the American billion (10^9) and trillion (10^{12}) are meant. The use of ppm, ppb and ppt is *only* permitted if they refer to mass/mass or volume/volume ratios; they should **not** be used for mass/volume ratios. The first time such a “unit” appears in an article, it should be indicated whether it refers to mass/mass or to volume/volume.

Appendix 3: Abbreviations and symbols that may be used without definition

Abbreviations and symbols should not be used in article titles. Please note that most abbreviations should only be used in combination with a value, or in structural formulae.

Abbreviations

| | |
|--|--|
| A, C, G, T | adenine, cytidine, guanine, thymine |
| Ac, OAc | acetyl, acetate |
| A/D | analog-to-digital |
| ADP, AMP, ATP, and similar nucleoside phosphates | adenosine 5'-di-, -mono-, triphosphate, etc. |
| a.c. | alternating current |
| amino acids | standard 3- and 1-letter codes |
| AU | absorbance units |
| BET | Brunauer—Emmett—Teller |
| b.p. | boiling point |
| Bu | butyl |
| cpm | counts per minute |
| CE | capillary electrophoresis |
| d, m, p, r, t (in nucleosides/nucleotides/nucleic acids) | deoxy, messenger, phosphate, recombinant/ribosomal, transfer |
| d.c. | direct current |
| DDD, DDT, DDE | di-, trichloro-bis(chlorophenyl)ethane, -ethylene |
| DEAE | diethylaminoethyl |
| DNA, DNase | deoxyribonucleic acid, deoxyribonuclease |
| Dns, dansyl | 5-dimethylaminonaphthalene-1-sulfonyl |

| | |
|-------------------------|--|
| DOPA | 3,4-dihydroxyphenylalanine |
| dpm | disintegrations per minute |
| EC | enzyme commission numbering system |
| EDTA | ethylenediaminetetraacetate, -acetic acid |
| equiv. | equivalent |
| Et | ethyl |
| FS | full scale |
| FSOT | fused-silica open tubular |
| FT | Fourier transform |
| GC, GLC, GSC | gas chromatography, gas-liquid chromatography, gas-solid chromatography |
| HP... | high-performance... |
| I.D. | internal diameter |
| IgG | immunoglobulin G |
| i.m. | intramuscular |
| i.p. | intra-peritoneal |
| IR | infrared |
| I.S. | internal standard |
| I.U. | international unit |
| i.v. | intravenous |
| LC | liquid chromatography |
| LD | lethal dose |
| Me | methyl |
| m.p. | melting point |
| MS | mass spectrometry |
| NAD, NADH (NADP, NADPH) | nicotinamide—adenine dinucleotide (phosphate) |
| NMR | nuclear magnetic resonance |
| O.D. | outer diameter |
| Ph | phenyl |
| Pr | propyl |
| PTFE | poly(tetrafluoroethylene) |
| RNA, RNase | ribonucleic acid, ribonuclease |
| RP... | reversed-phase.... |
| rpm | revolutions per minute |
| RSD | relative standard deviation (preferred over coefficient of variation) |
| SD | standard deviation |
| TLC | thin-layer chromatography |
| Tris | tris(hydroxymethyl)aminomethane |
| u | atomic mass units (reference to mass of ¹² C; preferred over a.m.u./amu: reference to mass of ¹⁶ O) |
| UV | ultraviolet |
| vol., v/v | volume, volume/volume |
| Vis | visible |
| WCOT | wall-coated open tubular |
| wt., w/w, m/m | mass, mass/mass |

Symbols

| | |
|---------------|--|
| <i>A</i> | peak area or absorbance |
| α | separation factor |
| <i>D</i> | diffusion coefficient |
| d_f | film thickness |
| d_p | particle diameter |
| ε | interparticle porosity or molar adsorptivity |
| <i>F</i> | mobile phase flow-rate |
| ΔG^0 | standard Gibbs free energy change |
| ΔH^0 | standard enthalpy change |
| <i>H</i> | plate height |
| <i>h</i> | reduced plate height |

| | |
|------------------|--|
| J | coupling constant |
| K | equilibrium constant |
| k | retention factor |
| K_c | distribution constant (preferred over partition coefficient) |
| L | length |
| λ | wavelength |
| M_r | (relative) molecular mass |
| μ | electrophoretic mobility |
| N | number of plates |
| n | number of determinations |
| η | viscosity |
| p | pressure or probability |
| P | relative pressure |
| $p\cdots$ | negative logarithm of... (as in pH, pI, pK _a) |
| r | relative retention or correlation coefficient |
| R | molar gas constant |
| R_F | retardation factor |
| R_M | $\log (1/R_F - 1)$ |
| R_s | resolution |
| ρ | density |
| ΔS^0 | standard entropy change |
| S/N | signal-to-noise ratio |
| T | temperature |
| t | time |
| t_0 | retention time of unretained compound |
| t_R (t'_R) | (adjusted) retention time |
| u | mobile phase velocity |
| V_0 | retention volume of unretained compound |
| V_R (V'_R) | (adjusted) retention volume |
| w_b | peak width at base |
| w_h | peak width at half height |

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